

# Fluorogenic Resveratrol-Confined Graphene Oxide For Economic and Rapid Detection Of Alzheimer's Disease

Xiao-Peng He,<sup>†</sup> Qiong Deng,<sup>†</sup> Liang Cai,<sup>†</sup> Chang-Zheng Wang,<sup>†,‡</sup> Yi Zang,<sup>\*,‡</sup> Jia Li,<sup>\*,‡</sup> Guo-Rong Chen,<sup>\*,†</sup> and He Tian<sup>†</sup>

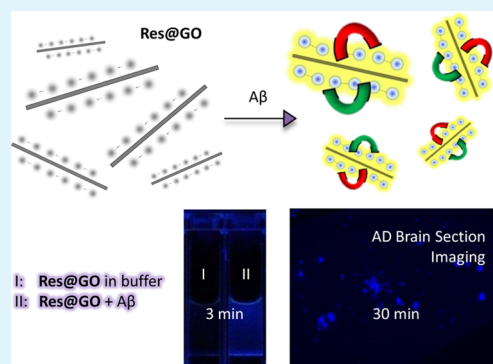
<sup>†</sup>Key Laboratory for Advanced Materials & Institute of Fine Chemicals, East China University of Science and Technology, 130 Meilong Road, Shanghai 200237, P. R. China

<sup>‡</sup>National Center for Drug Screening, State Key Laboratory of Drug Research, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, 189 Guo Shoujing Road, Shanghai 201203, P. R. China

## S Supporting Information

**ABSTRACT:** Developing an effective means for the real-time probing of amyloid  $\beta$  ( $A\beta$ ) that is closely implicated in Alzheimer's disease (AD) could help better understand and monitor the disease. Here we describe an economic approach based on the simple composition of a natural product, resveratrol (Res), with graphene oxide (GO) for the rapid, fluorogenic recognition of  $A\beta$ . The Res@GO composite has proved specific for  $A\beta$  over a range of proteins and ions, and could sensitively capture both  $A\beta$  monomers and fibers in a physiological buffer solution within only 3 min. The composite can also fluorescently image amyloid deposits in a mouse brain section within 30 min. This new protocol is much cheaper and more timesaving than the conventional immunofluorescence staining technique employed clinically, providing an economic tool for the concise detection of AD.

**KEYWORDS:** graphene, Alzheimer's disease, resveratrol, amyloid  $\beta$ , fluorogenic, detection



Alzheimer's disease (AD), the most common form of dementia, suffers tens of millions of people per year and is estimated to impact one out of 85 people globally by 2050.<sup>1,2</sup> Unfortunately, the pathogenesis and progression of AD have not been well understood, marking the disease among the most costly social burdens in developed countries.<sup>3,4</sup> Furthermore, the fact that there are currently no treatments available that cure or reverse the progression of the disease implicates that effective means for the better dissection of AD is both challenging and highly desirable.

Amyloid  $\beta$  ( $A\beta$ ) that is truncated successively by  $\beta$  and  $\gamma$  secretases from the amyloid precursor protein<sup>5</sup> is tightly associated with the pathology of AD; it is the main component of amyloid plaques found in the brains of AD patients.<sup>6</sup>  $A\beta$  can form not only tangles of regularly organized fibrillar aggregates (the  $A\beta$  fibrils) in the brain but also highly toxic soluble oligomers that are believed to boost the development of AD.<sup>5,7,8</sup> As a result, sensitive monitoring of  $A\beta$  of various forms under physiological conditions is of paramount importance towards the elucidation of AD. However, current techniques (e.g., immunofluorescence staining) used in laboratories and hospitals for detection of  $A\beta$  in solution and brain sections is expensive and onerous, and thus alternative tools that address these issues should be developed.

Graphene oxide (GO),<sup>9</sup> because of its promising optical properties, good water solubility, low toxicity, and cost-effectiveness, has become a material of choice for the

development of bioprobes.<sup>10–17</sup> GO has also proved to be a universal quencher of fluorescence (FL) dyes through the fluorescence resonance energy transfer (FRET) effect.<sup>12,18</sup> A recent study demonstrates that, comparing to carbon nanotubes and gold nanoparticles, GO is more efficient for DNA analyses.<sup>19</sup> We also exemplify that GO may serve as a potent and economic medium in the fabrication of optical or electrochemical composite bioprobes.<sup>20,21</sup>

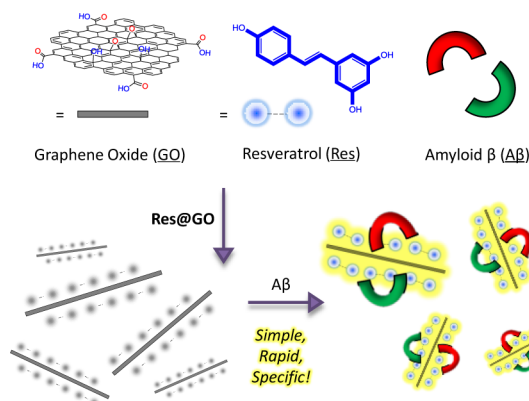
Given the said interesting aspects of GO, we sought to fabricate a GO-based fluorogenic probe for the rapid and cost-effective determination of  $A\beta$ . Resveratrol (Res), a natural product widely found in red wine, has shown neuroprotective effects<sup>22</sup> and could bind specifically with  $A\beta$ .<sup>23,24</sup> On the basis of its intrinsic FL, we devised a new probe composite that utilizes Res as both the signaling and recognition molecule. Scheme 1 depicts the design rationale behind which to construct the fluorogenic Res@GO (GO confined with Res). Stacking of the fluorescent Res to the surface of GO by, probably,  $\pi$ -interactions generates a fluorogenic composite that quenches the FL of Res via FRET. While in complexation with an  $A\beta$ , the composite will fluoresce because of impeded energy transfer between GO and the photoexcited Res specifically encapsulated by the peptide.

**Received:** February 21, 2014

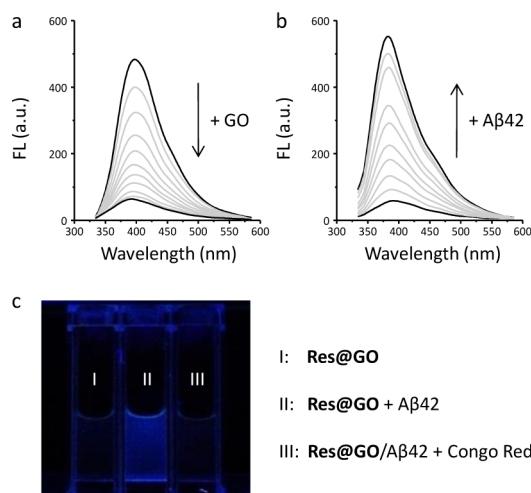
**Accepted:** April 4, 2014

**Published:** April 4, 2014

**Scheme 1. Structure of Resveratrol (Res) and Graphene Oxide (GO), and Cartoon Depicting the Simple Construction of Res@GO for the Rapid Fluorogenic Probing of Amyloid  $\beta$  ( $A\beta$ )**



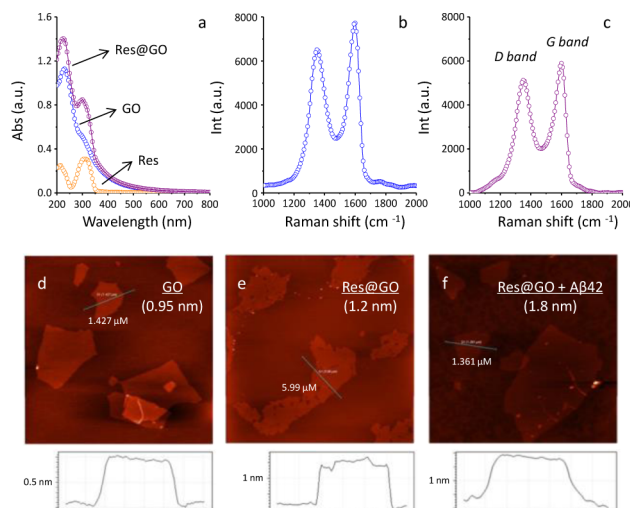
To begin with, the quenching effect of GO for Res (20  $\mu\text{M}$ ) was tested. As shown in Figure 1a, increasing GO (0–50  $\mu\text{g}/$



**Figure 1.** Fluorescence (FL) titration spectra of (a) resveratrol (Res, 20  $\mu\text{M}$ ) with increasing graphene oxide (GO, 0–50  $\mu\text{g}/\text{mL}$ ), and (b) Res@GO with increasing monomeric amyloid  $\beta$ 42 ( $A\beta$ 42, 0–200  $\mu\text{M}$ ) in phosphate buffered saline (0.2 M, pH 7.4) with excitation at 310 nm. (c) Photographed FL change of Res@GO (I) in the presence of  $A\beta$ 42 (II) and then congo red (III) excited with a portable UV lamp.

mL) led to gradual FL (excited at 310 nm) decrease of the natural product with a maximum quenching rate of 88.4% ( $F/F_0$ , where  $F$  is the quenched FL and  $F_0$  the original FL). This is probably caused by the FRET effect between the photo-excited Res and GO, the energy acceptor, upon composition.<sup>12,18,20</sup> Then, we observed that addition of increasing monomeric  $A\beta$ 42 to the composited Res@GO led to rapid (3 min incubation) and sharp FL recovery of the composite in a concentration-dependent manner (Figure 1b). Figure 1c illustrates the fluorogenic response of the composite to  $A\beta$ 42 (I to II), whereas further addition of congo red (a known strong  $A\beta$  binder)<sup>25</sup> competitively quenched the FL (III). This means that the FL “off–on” detection is reversible.

To corroborate the stacking between Res and GO, a variety of analytical techniques were employed. UV–vis (Figure 2a) and Fourier transmission infrared (FTIR) spectra (see Figure

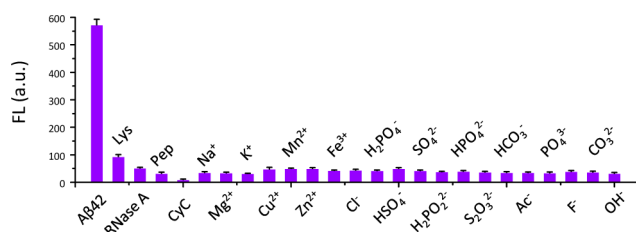


**Figure 2.** (a) UV–vis spectra of resveratrol (Res), graphene oxide (GO), and Res@GO. Raman spectra of (b) GO, and (c) Res@GO. Atomic force microscope image of (d) GO, (e) Res@GO, and (f) Res@GO in complex with amyloid  $\beta$ 42 ( $A\beta$ 42).

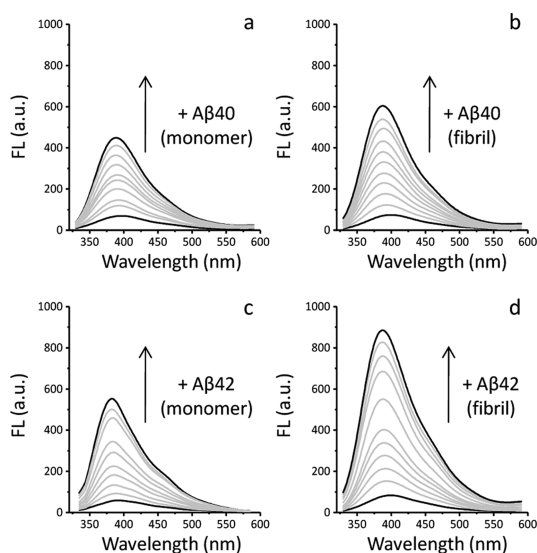
S2 in the Supporting Information) first showed that Res@GO displays peaks characteristic of both the compound and the carbon flake. Typical D band (1355  $\text{cm}^{-1}$ ) and G band (1600  $\text{cm}^{-1}$ ) curves were recorded in the Raman spectrum of GO with an  $I_D/I_G$  (where  $I$  is intensity) ratio of 0.83 (Figure 2b). In contrast, the  $I_D/I_G$  ratio of Res@GO increased to 0.89 (Figure 2c), suggesting an increased  $\text{sp}^2$ -hybridization disorder of GO composited with the aromatic Res.<sup>20,21</sup> In addition, the atomic force microscope (AFM) image of bare GO (Figure 2d) showed that the height of which is 0.95 nm, whereas stacking of Res to GO increased the height to 1.2 nm (Figure 2e). Complexation of Res@GO with  $A\beta$ 42 further thickened the composite to 1.8 nm.

These data, together with the FL titration results shown in Figure 1, suggest that the stacked Res@GO composite may serve as a fluorogenic probe that can rapidly adhere  $A\beta$ 42 onto its Res-confined surface, leading to FL enhancement due to the encapsulation of the Res cluster by the peptide. The specificity of Res@GO for  $A\beta$  was interrogated using a panel of physiological proteins including ribonuclease A (RNaseA), lysosome (Lys), pepsin (pep), and cytochrome c (CyC), and a variety of cations and anions as shown in Figure 3. We determined that all these species hardly fluctuated the FL of the composite, which manifests the good specificity of the concisely constructed Res@GO for  $A\beta$ 42.

Next, as it has been reported that  $A\beta$  with 40 and 42 residues are the main components to form insoluble amyloid fibrils that eventually induce necrosis of neuronal cells,<sup>5–8</sup> we tested the recognition of Res@GO using both monomeric (m-) and fibril (-)  $A\beta$ 40/ $A\beta$ 42. The  $A\beta$  fibrils were produced as indicated in a previous study.<sup>14</sup> As shown in Figure 4, although its response to f- $A\beta$ 40 (Figure 4b) is slightly stronger than that to m- $A\beta$ 40 (Figure 4a), Res@GO showed evidently more increased FL with f- $A\beta$ 42 (Figure 4d) than with m- $A\beta$ 42 (Figure 4c). This suggests that the higher binding affinity of Res with the  $A\beta$  fibrils than the monomers<sup>23</sup> could be accurately interpreted by the composite. Furthermore, good linear ranges as regards the FL alternation in response to the analytes were produced by the composite (see Figure S2 in the Supporting Information); the



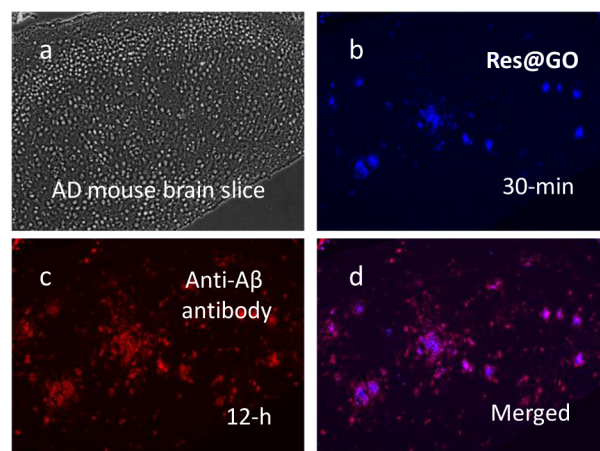
**Figure 3.** FL change of Res@GO (20  $\mu\text{M}$  of resveratrol with 50  $\mu\text{g}/\text{mL}$  graphene oxide) in the presence of A $\beta$ 42 (200  $\mu\text{M}$ ) or a variety of physiological proteins (200  $\mu\text{M}$ ) including ribonuclease A (RNase A), lysosome (Lys), pepsin (pep), and cytochrome c (CyC), and ions (10 mM) in phosphate buffered saline (0.2 M, pH 7.4) with excitation at 310 nm.



**Figure 4.** Fluorescence (FL) titration spectra of Res@GO (20  $\mu\text{M}$  of resveratrol with 50  $\mu\text{g}/\text{mL}$  graphene oxide) with increasing (a) A $\beta$ 40 monomer (0–200  $\mu\text{M}$ ), (b) A $\beta$ 40 fibril (0–200  $\mu\text{M}$ ), (c) A $\beta$ 42 monomer (0–200  $\mu\text{M}$ ), and (d) A $\beta$ 42 fibril (0–120  $\mu\text{M}$ ) in phosphate buffered saline (0.2 M, pH 7.4) with excitation at 310 nm.

limits of detection for m-A $\beta$ 40, m-A $\beta$ 42, f-A $\beta$ 40, and f-A $\beta$ 42 were determined to be 277, 219, 270, and 120 nM, respectively.

Having demonstrated the good sensitivity and selectivity of Res@GO for A $\beta$ , we attempted its applicability in imaging amyloid plaques formed in a brain section. Section collected from the brain (Figure 5a) of a transgenic mouse carrying APP<sup>swe</sup>/PS1<sup>1dE9</sup> mutations (APP/PS1 mouse, a kind gift given by prof. Haiyan Zhang of Shanghai Institute of Materia Medica)<sup>26</sup> was used to be imaged by the composite or by an anti-A $\beta$  antibody as a positive control. As the composite will remain at its quenching state unless it binds with A $\beta$ , the conventional rinsing step after treatment with Res@GO was envisioned to be unnecessary. Indeed, we observed that after incubation of the section with Res@GO for only 30 min, clear FL imaging of the plaques was recorded with almost no background FL observed (Figure 5b). The imaged area was well-correlated to that immunofluorescently stained by the specific anti-A $\beta$  antibody (Figure 5c, d). The composite also showed little toxicity to a human normal cell line (human embryonic kidney 293, HEK293), even with a 5-fold elevated concentration used for imaging (see Figure S3 in the Supporting Information).<sup>27</sup>



**Figure 5.** (a) Bright-field image of a mouse brain section with amyloid plaques; fluorescence imaging of the plaques using (b) Res@GO (wash-free) and (c) anti-A $\beta$  antibody; (d) the merged fluorescence staining.

We note that, for the brain section imaging, the Res@GO composite simply developed here is much more advantageous than the conventional immunofluorescence staining in terms of the following: (1) whereas only  $\sim 30$  min was required from the treatment of the brain section with Res@GO to FL imaging, approximately 12 h was needed for the antibody staining (which includes the onerous blocking processes, primary/secondary antibody labeling, and repeated washing to rinse off residual species); (2) the cost of the conventional technique was estimated to be  $\sim \text{¥}1500$  per 100 slices (roughly taking into account the price of antibodies), whereas that of the present protocol  $\sim \text{¥}0.85$  per 100 slices (roughly taking into account the price of GO and Res; details for cost estimation are shown in the Supporting Information). As a consequence, Res@GO could become a potentially economic and timesaving tool for both laboratorial and clinical uses.

In summary, we have developed a succinct fluorogenic Res@GO composite for the rapid and cost-effective detection of A $\beta$  in solution as well as brain section. This low-cost material might hold the promise to replace antibody-based immunofluorescence staining of clinical samples and serve as a reliable probe for the fast determination of A $\beta$  of various forms. This study also offers valuable insights into the concise dissection of AD.

## ■ ASSOCIATED CONTENT

### Supporting Information

Experimental section and additional figures. This material is available free of charge via the Internet at <http://pubs.acs.org/>.

## ■ AUTHOR INFORMATION

### Corresponding Authors

\*E-mail: yzang@mail.shcnc.ac.cn.

\*E-mail: jli@mail.shcnc.ac.cn.

\*E-mail: mrs\_guorongchen@ecust.ac.cn.

### Notes

The authors declare no competing financial interest.

## ■ ACKNOWLEDGMENTS

We thank the 973 project (2013CB733700), the National Science Fund for Distinguished Young Scholars (81125023), the National Natural Science Foundation of China (21176076,

21202045 and 81173033), and the Key Project of Shanghai Science and Technology Commission (13NM1400900).

## REFERENCES

- (1) Brookmeyer, R.; Gray, S.; Kawas, C. Projections Of Alzheimer's Disease In The United States And The Public Health Impact Of Delaying Disease Onset. *Am. J. Public Health* **1998**, *88*, 1337–1342.
- (2) Brookmeyer, R.; Johnson, E.; Ziegler-Graham, K.; Arrighi, M. H. Forecasting The Global Burden Of Alzheimer's Disease. *Alzheimer's Dementia* **2007**, *3*, 186–191.
- (3) Meek, P. D.; McKeithan, K.; Schumock, G. T. Economic Considerations In Alzheimer's Disease. *Pharmacotherapy* **1998**, *18*, 68–73.
- (4) Bonin-Guillaume, S.; Zekry, D.; Giacobini, E.; Gold, G.; Michel, J.-P. The Economical Impact Of Dementia. *Presse Med.* **2005**, *34*, 35–41.
- (5) Hartmann, T.; Bieger, S. C.; Brühl, B.; Tienari, P. J.; Ida, N.; Allsop, D.; Roberts, G. W.; Masters, C. L.; Dotti, C. G.; Unsicker, K.; Beyreuther, K. Distinct Sites Of Intracellular Production For Alzheimer's Disease A $\beta$ 40/42 Amyloid Peptides. *Nat. Med.* **1997**, *3*, 1016–1020.
- (6) Eisenberg, D.; Jucker, M. The Amyloid State Of Proteins In Human Disease. *Cell* **2012**, *148*, 1188–1203.
- (7) Prelli, F.; Castaño, E.; Glenner, G. G.; Frangione, B. Differences Between Vascular And Plaque Core Amyloid In Alzheimer's Disease. *J. Neurochem.* **1988**, *51*, 648–651.
- (8) Zhao, L. N.; Long, H. W.; Mu, Y.; Chew, L. Y. The Toxicity Of Amyloid  $\beta$  Oligomers. *Int. J. Mol. Sci.* **2012**, *13*, 7303–7327.
- (9) Novoselov, K. S.; Geim, A. K.; Morozov, S. V.; Jiang, D.; Zhang, Y.; Dubonos, S. V.; Grigorieva, I. V.; Firsov, A. A. Electric Field Effect In Atomically Thin Carbon Films. *Science* **2004**, *306*, 666–669.
- (10) Chen, D.; Feng, H.; Li, J. Graphene Oxide: Preparation, Functionalization, And Electrochemical Applications. *Chem. Rev.* **2012**, *112*, 6027–6053.
- (11) Georgakilas, V.; Otyepka, M.; Bourlinos, A. B.; Chandra, V.; Kim, N.; Kemp, K. C.; Hobza, P.; Zboril, R.; Kim, K. S. Functionalization Of Graphene: Covalent And Non-Covalent Approaches, Derivatives And Applications. *Chem. Rev.* **2012**, *112*, 6156–6214.
- (12) Morales-Narváez, E.; Merkoçi, A. Graphene Oxide As An Optical Biosensing Platform. *Adv. Mater.* **2012**, *24*, 3298–3308.
- (13) Feng, L.; Wu, L.; Qu, X. New Horizons For Diagnostics And Therapeutics Applications Of Graphene And Graphene Oxide. *Adv. Mater.* **2013**, *25*, 168–186.
- (14) Li, M.; Yang, X.; Ren, J.; Qu, K.; Qu, X. Using Graphene Oxide High Near-Infrared Absorbance For Photothermal Treatment Of Alzheimer's Disease. *Adv. Mater.* **2012**, *24*, 1722–1728.
- (15) Wu, J.-J.; Zhao, W.-W.; Song, S.; Fan, C.; Chen, H.-Y. Functional Nanoprobes For Ultrasensitive Detection Of Biomolecules: An Update. *Chem. Soc. Rev.* **2014**, *43*, 1601–1611.
- (16) Pei, H.; Li, J.; Lv, M.; Wang, J.; Gao, J.; Lu, J.; Li, Y.; Huang, Q.; Hu, J.; Fan, C. A Graphene-Based Sensor Array For High-Precision And Adaptive Target Identification With Ensemble Aptamers. *J. Am. Chem. Soc.* **2012**, *134*, 13843–13849.
- (17) Liu, X.; Wang, F.; Aizen, R.; Yehezkeli, O.; Willner, I. Graphene Oxide/Nucleic-Acid-Stabilized Silver Nanoclusters: Functional Hybrid Materials For Optical Aptamer Sensing And Multiplexed Analysis Of Pathogenic DNAs. *J. Am. Chem. Soc.* **2013**, *135*, 11832–11839.
- (18) Kim, J.; Cote, L. J.; Kim, F.; Huang, J. Visualizing Graphene Based Sheets By Fluorescence Quenching. *J. Am. Chem. Soc.* **2010**, *132*, 260–267.
- (19) Fan, L.; Pei, H.; Wang, L.; Lu, J.; Gao, J.; Jiang, B.; Zhao, X.; Fan, C. Nanomaterial-Based Fluorescent DNA Analysis: A Comparative Study Of The Quenching Effects Of Graphene Oxide, Carbon Nanotubes, and Gold Nanoparticles. *Adv. Funct. Mater.* **2013**, *23*, 4140–4148.
- (20) Zhang, H.-L.; Wei, X.-L.; Zang, Y.; Cao, J.-Y.; Liu, S.; He, X.-P.; Chen, Q.; Long, Y.-T.; Li, J.; Chen, G.-R.; Chen, K. Fluorogenic Probing Of Specific Recognitions Between Sugar Ligands And Glycoprotein Receptors On Cancer Cells By An Economic Graphene Nanocomposite. *Adv. Mater.* **2013**, *25*, 4097–4101.
- (21) Li, Z.; Deng, S.-S.; Zang, Y.; Gu, Z.; He, X.-P.; Chen, G.-R.; Chen, K.; James, T. D.; Li, J.; Long, Y.-T. Capturing Intercellular Sugar-Mediated Ligand-Receptor Recognitions Via A Simple Yet Highly Biospecific Interfacial System. *Sci. Rep.* **2013**, *3*, 2293.
- (22) Vingtdoux, V.; Dreses-Werringloer, U.; Zhao, H.; Davies, P.; Marambaud, P. Therapeutic Potential Of Resveratrol In Alzheimer's Disease. *BMC Neurosci.* **2008**, *9*, S6.
- (23) Ge, J.-F.; Qiao, J.-P.; Qi, C.-C.; Wang, C.-W.; Zhou, J.-N. The Binding Of Resveratrol To Monomer And Fibril Amyloid Beta. *Neurochem. Int.* **2012**, *61*, 1192–1201.
- (24) Ahn, J. S.; Lee, J.-H.; Kim, J.-H.; Paik, S. R. Novel Method For Quantitative Determination Of Amyloid Fibrils Of  $\alpha$ -Synuclein And Amyloid  $\beta$ /A4 Protein By Using Resveratrol. *Anal. Biochem.* **2007**, *367*, 259–265.
- (25) Howie, A. J.; Brewer, D. B. Optical Properties Of Amyloid Stained By Congo Red: History And Mechanisms. *Micron* **2009**, *40*, 285–301.
- (26) Zhang, B.; Tang, X. C.; Zhang, H. Y. Alternations Of Central Insulin-Like Growth Factor-1 Sensitivity In APP/PS1 Transgenic Mice And Neuronal Models. *J. Neurosci. Res.* **2013**, *91*, 717–725.
- (27) Liao, K.-H.; Lin, Y.-S.; Macosko, C. W.; Haynes, C. L. Cytotoxicity Of Graphene Oxide And Graphene In Human Erythrocytes And Skin Fibroblasts. *ACS Appl. Mater. Interfaces* **2011**, *3*, 2607–2615.